



WHAT'S LIVING IN YOUR ACTIVATED SLUDGE

Presenter:

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Outline

» Microbial Identification

- Culturing
- Microscope
- DNA Testing
- DNA Applications

» Case Study: Ho-Chunk WWTF

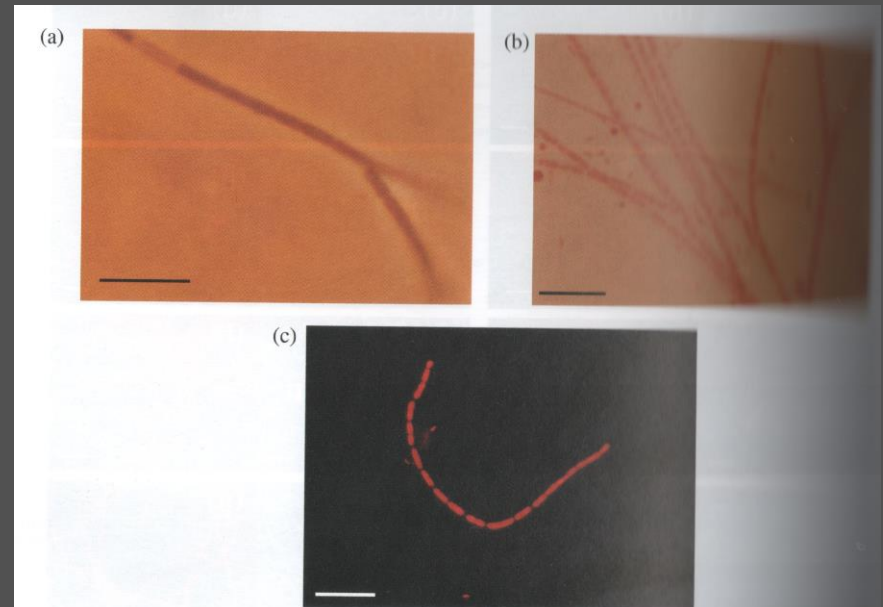
Moving Beyond Culture ID

- » Less than 1% of bacteria will grow in culture
- » Requires days



Moving Beyond the Microscope

- » Many bacteria cannot be identified visually
- » Multiple species look identical
- » Some species morph between shape based on environmental conditions



Example DNA Data

>594682

TACGGAGGATCCAAGCGTTATCCGGATTTATTGGGTTTAAAGGGTACGTAGGCGGACCCG
TCAGTCAGTGGTGAAAGTTTGCAGCTTAACTGTAAAATTGCCATTGAAACTACGGGTCTT
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>594511

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>594448

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GGAACACCCGTAGCGAAGGCGGCATTCTGGGCCGGTACTGACGCTGAGGAGCGAAAGCGT
GGGGAGCGAACAGG

Applications

- » Drinking water
- » Surface Waters
 - Source tracking of fecal pollution
- » Remediation
 - Track populations of microbial degraders
- » Air quality monitoring
- » Food and beverage
 - Safety, quality, longevity

Applications

» Wastewater process trouble-shooting

- Loss of nitrification
- Loss of Bio-P
- Poor digester gas production
- Pathogens in biosolids or effluent
- Source tracking of fecal pollution
- Bulking
- Foaming

Filaments

- » Microscope identification
- » >1 species present
- » Usually beneficial
- » Bulking: Too many filaments
- » Selectors
- » Diverse, complex causes



Foamers

» Foam

- Hydrophobic particles (bacteria)
- Surfactants (excreted by bacteria)
- Bubbles

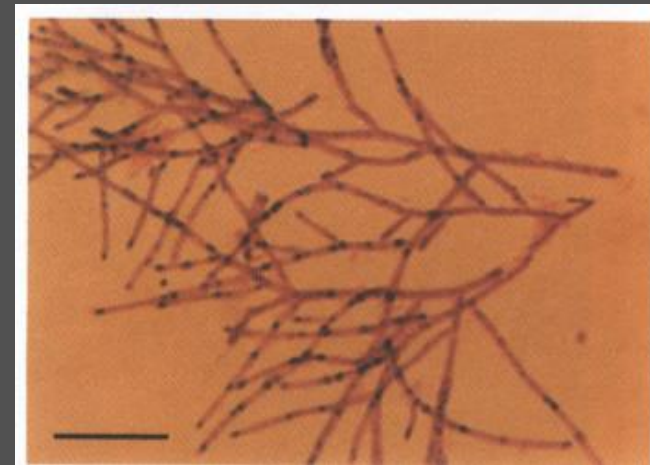
» Nocardia

» Microthrix

» Low abundance high impact

» Not necessarily filamentous

» Hard to get rid of it!



Ho-Chunk WWTF DNA Data



Ho-Chunk Village WWTF

» Sequencing Batch Reactor facility

» Design Conditions

- Flow – 0.30 mgd
- BOD – 1,270 lbs/day

» Current Loadings

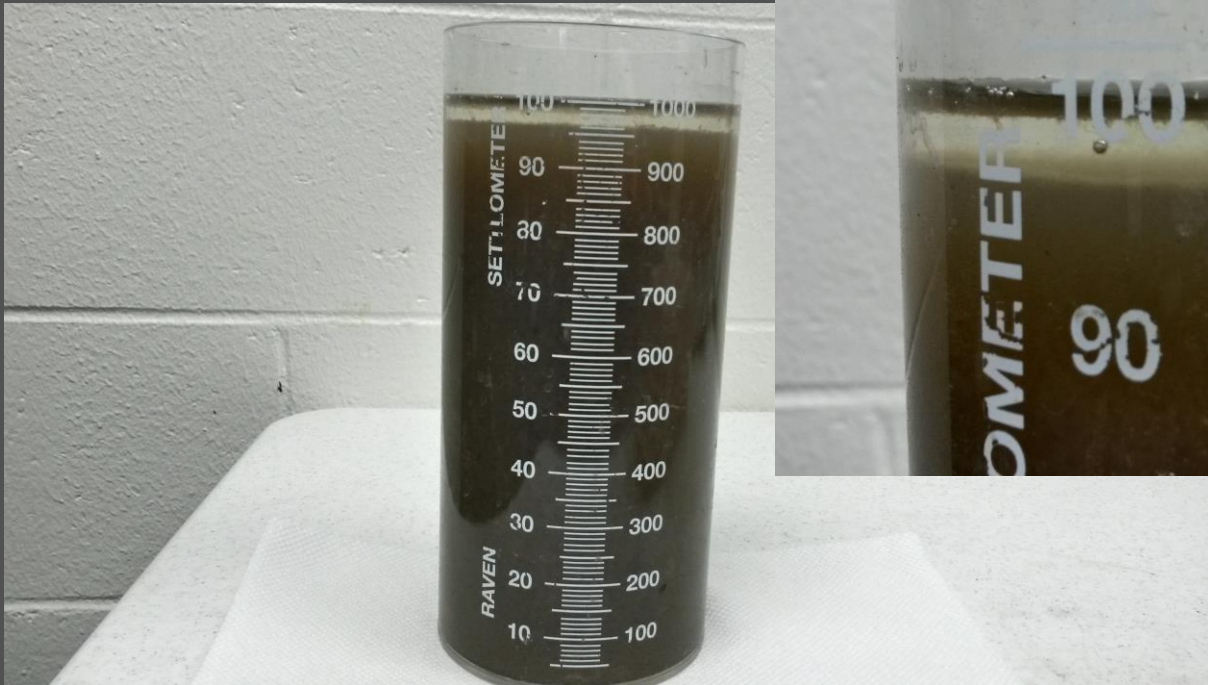
- Flow – 0.06 mgd
- BOD – 350 lbs/day

» Effluent Limits

- BOD < 50 mg/L
- Total N < 10 mg/L (need to nitrify and denitrify)

Ho-Chunk Village WWTF

- » Poor settling during cold weather
- » High SVI has lead to elevated effluent TSS



DNA Testing Program

- » Samples collected weekly April – August 2014
- » All testing was done on one reactor (SBR #4)
- » Total of 21 samples were frozen and shipped in batches for bacterial assessment

Sample Results - General

- » Nitrogen Removers (13 of 27 detected)
- » Filaments (9 of 18 detected)
- » Foaming (6 of 9 detected)
- » Results are in % of total bacteria in a known volume

Filamentous Bacteria Results

Date	4/9/2014	4/15/2014	4/22/2014	4/29/2014	5/6/2014	5/17/2014	5/20/2014	5/27/2014	6/3/2014	6/10/2014	6/17/2014
Thiothrix	0.01%	0.01%	0.01%	0.02%	0.01%	0.01%	0.01%	0.01%	0.01%	0.01%	0.01%
Acinetobacter	0.09%	0.10%	0.03%	0.04%	0.03%	0.02%	0.02%	0.04%	0.05%	0.07%	0.05%
Microthrix	0.14%	0.48%	0.41%	0.31%	0.06%	0.48%	1.74%	1.04%	0.44%	0.69%	1.86%
Runella	0.07%	0.11%	0.11%	0.05%	0.10%	0.08%	0.08%	0.08%	0.11%	0.08%	0.03%
Gordonia	0.01%	0.02%	0.01%	0.01%	0.01%	0.01%	0.02%	0.02%	0.01%	0.01%	0.01%
Haliscomenobacter	0.04%	0.01%	0.03%	0.03%	0.05%	0.03%	0.04%	0.02%	0.03%	0.05%	0.04%
Leptolinea	0.02%	0.02%	0.01%	0.02%	0.02%	0.02%	0.03%	0.02%	0.03%	0.02%	0.03%
Caldilinea	0.25%	0.29%	0.26%	0.28%	0.13%	0.15%	0.27%	0.16%	0.10%	0.11%	0.07%
Kouleoithrix	0.25%	0.37%	0.36%	0.34%	0.60%	0.38%	0.63%	0.74%	0.82%	0.73%	0.80%

Date	6/24/2014	7/1/2014	7/8/2014	7/15/2014	7/22/2014	7/29/2014	8/7/2014	8/12/2014	8/19/2014	8/26/2014
Thiothrix	0.00%	0.01%	0.00%	0.00%	0.00%	0.00%	0.00%	0.01%	0.00%	0.00%
Acinetobacter	0.08%	0.06%	0.05%	0.04%	0.04%	0.03%	0.06%	0.08%	0.08%	0.08%
Microthrix	1.75%	0.37%	1.31%	0.96%	0.14%	0.30%	0.15%	0.04%	0.02%	0.00%
Runella	0.11%	0.11%	0.10%	0.07%	0.05%	0.04%	0.03%	0.03%	0.03%	0.02%
Gordonia	0.05%	0.10%	0.03%	0.04%	0.01%	0.01%	0.02%	0.01%	0.01%	0.01%
Haliscomenobacter	0.02%	0.03%	0.05%	0.07%	0.09%	0.08%	0.07%	0.19%	0.12%	0.09%
Leptolinea	0.00%	0.00%	0.00%	0.00%	0.00%	0.00%	0.00%	0.00%	0.00%	0.00%
Caldilinea	0.53%	0.33%	0.34%	0.32%	0.23%	0.22%	0.24%	0.24%	0.22%	0.21%
Kouleoithrix	1.34%	1.92%	1.67%	2.14%	1.54%	1.91%	1.76%	1.84%	1.86%	1.72%

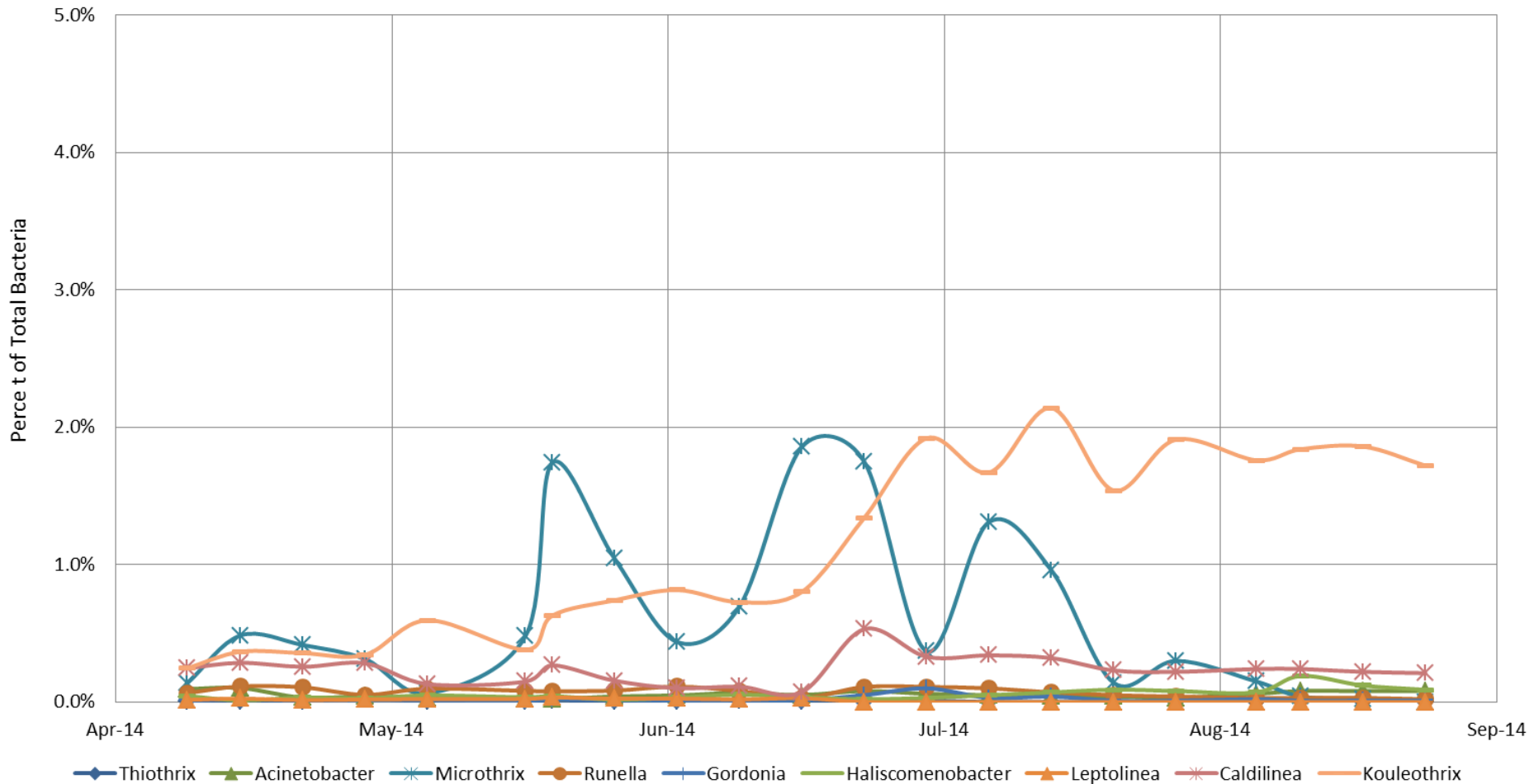
Foaming Bacteria Results

Date	4/9/2014	4/15/2014	4/22/2014	4/29/2014	5/6/2014	5/17/2014	5/20/2014	5/27/2014	6/3/2014	6/10/2014	6/17/2014
Acidimicrobiales	1.69%	2.69%	1.90%	1.62%	0.88%	1.28%	3.17%	1.78%	1.32%	1.36%	2.74%
Microthrix	0.14%	0.48%	0.41%	0.31%	0.06%	0.48%	1.74%	1.04%	0.44%	0.69%	1.86%
Rhodococcus	0.01%	0.01%	0.01%	0.00%	0.01%	0.01%	0.01%	0.01%	0.01%	0.01%	0.00%
Actinomycetales	1.25%	1.35%	1.01%	1.07%	1.10%	0.79%	1.25%	0.45%	0.51%	0.73%	0.55%
Gordonia	0.01%	0.02%	0.01%	0.01%	0.01%	0.01%	0.02%	0.02%	0.01%	0.01%	0.01%
Mycobacterium	0.03%	0.03%	0.03%	0.03%	0.03%	0.03%	0.04%	0.02%	0.02%	0.04%	0.02%

Date	6/24/2014	7/1/2014	7/8/2014	7/15/2014	7/22/2014	7/29/2014	8/7/2014	8/12/2014	8/19/2014	8/26/2014
Acidimicrobiales	2.02%	1.39%	1.71%	1.71%	1.52%	1.61%	2.42%	1.66%	1.09%	0.70%
Microthrix	1.75%	0.37%	1.31%	0.96%	0.14%	0.30%	0.15%	0.04%	0.02%	0.00%
Rhodococcus	0.09%	0.05%	0.04%	0.03%	0.02%	0.03%	0.03%	0.02%	0.02%	0.01%
Actinomycetales	1.72%	1.37%	1.02%	1.00%	0.69%	0.63%	0.82%	0.61%	0.34%	0.30%
Gordonia	0.05%	0.10%	0.03%	0.04%	0.01%	0.01%	0.02%	0.01%	0.01%	0.01%
Mycobacterium	0.11%	0.12%	0.08%	0.10%	0.07%	0.07%	0.05%	0.03%	0.03%	0.02%

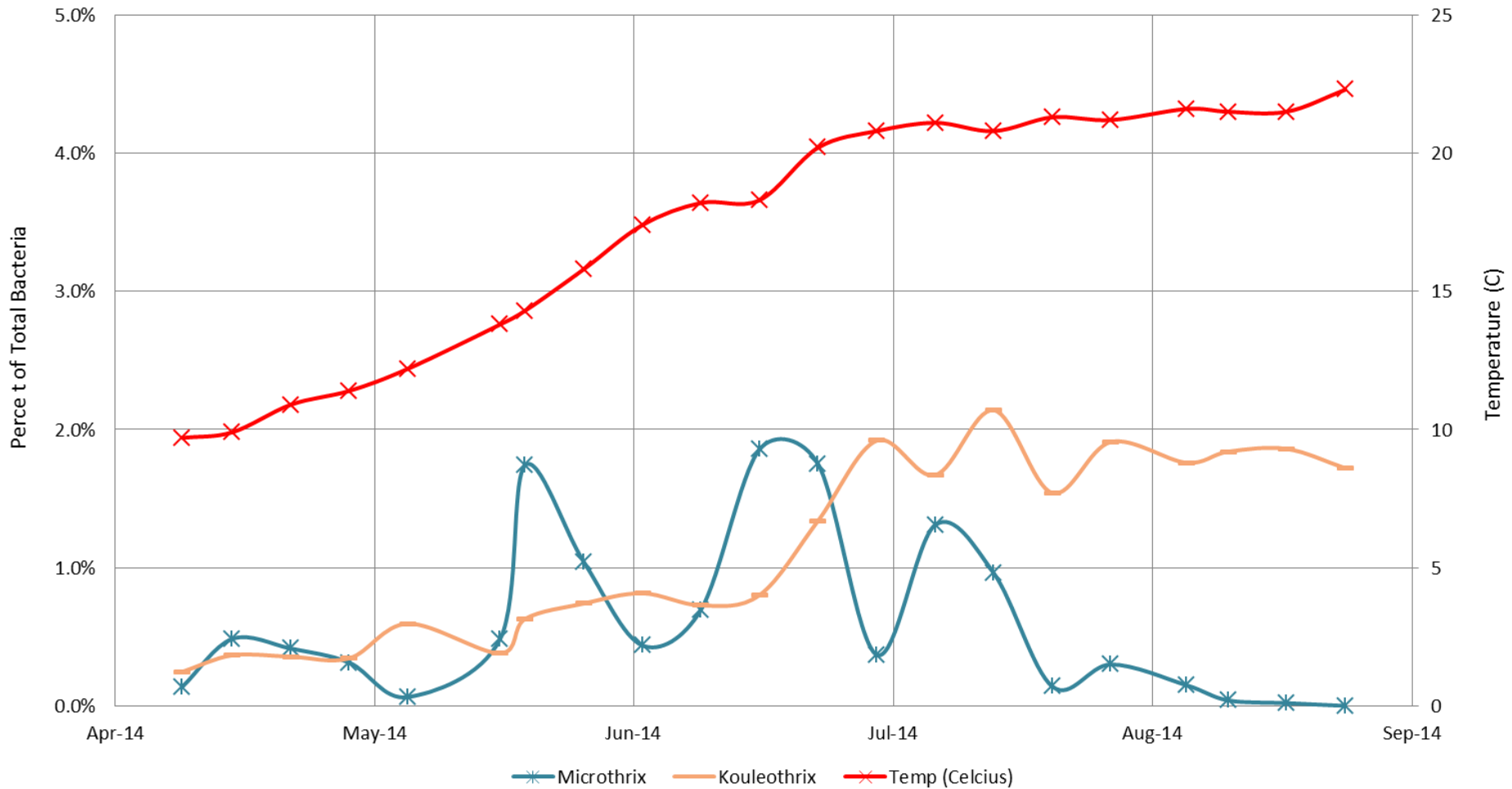
Filamentous Bacteria Results

Filamentous Bacteria

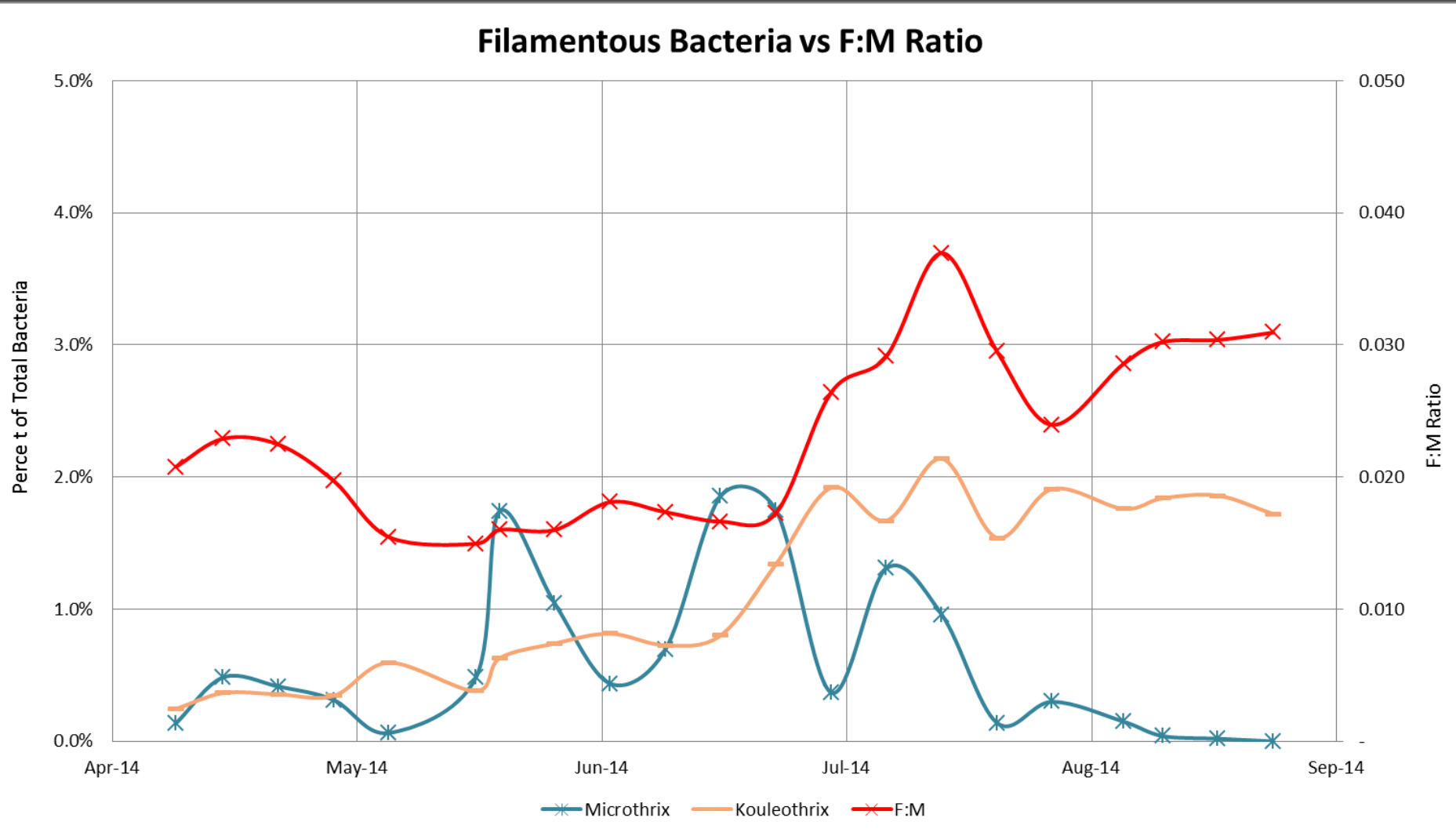


Filamentous Bacteria Results

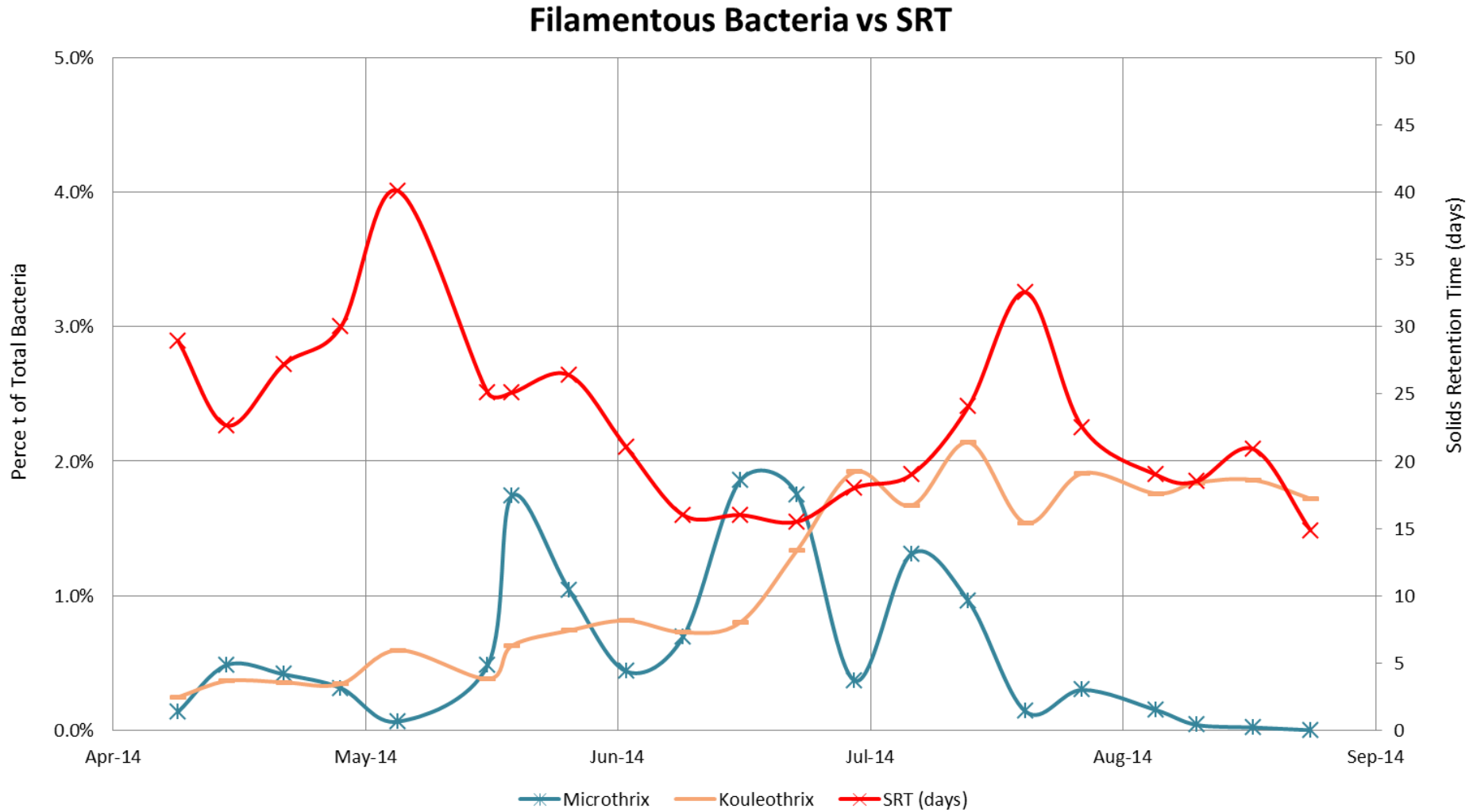
Filamentous Bacteria vs Temperature



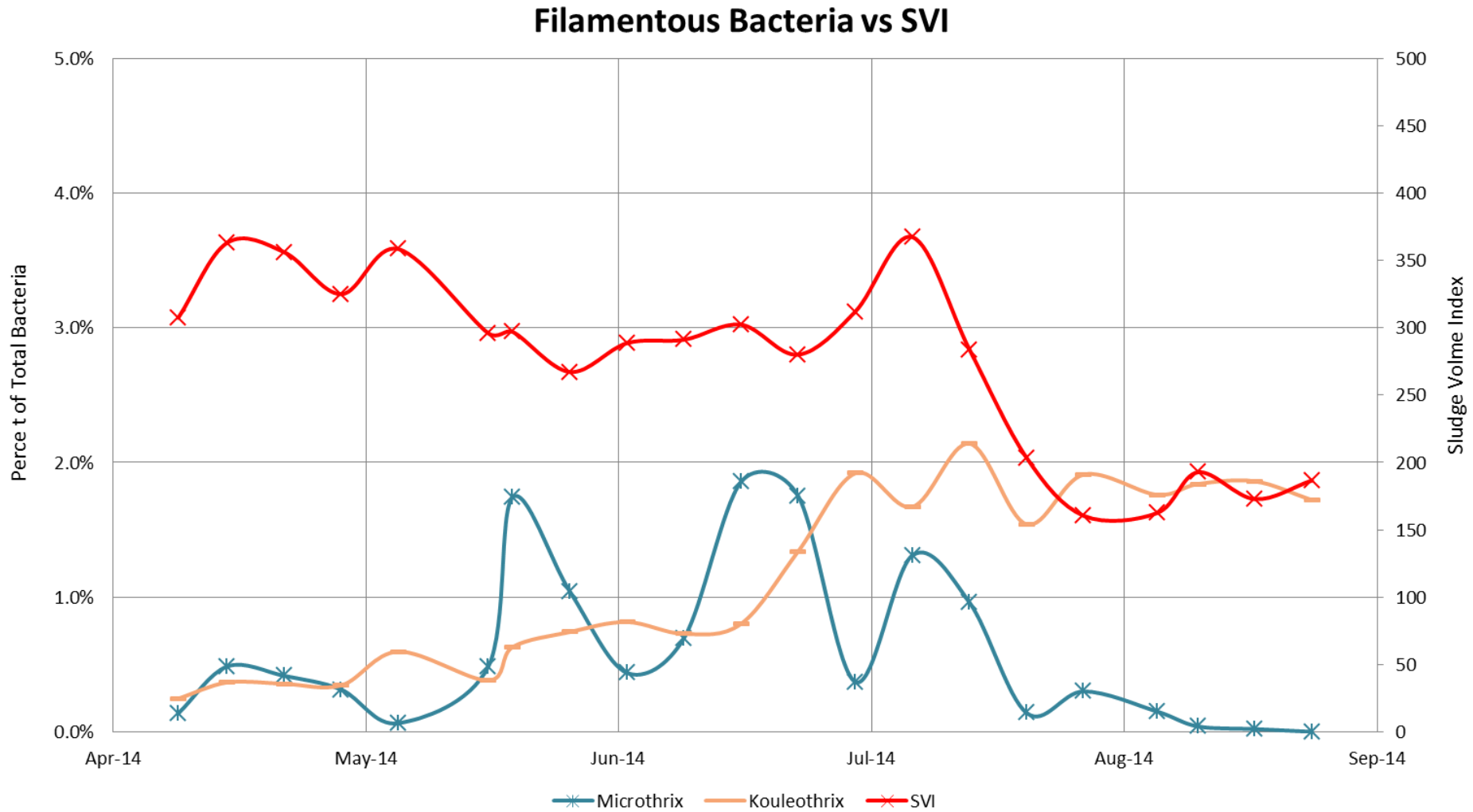
Filamentous Bacteria Results



Filamentous Bacteria Results

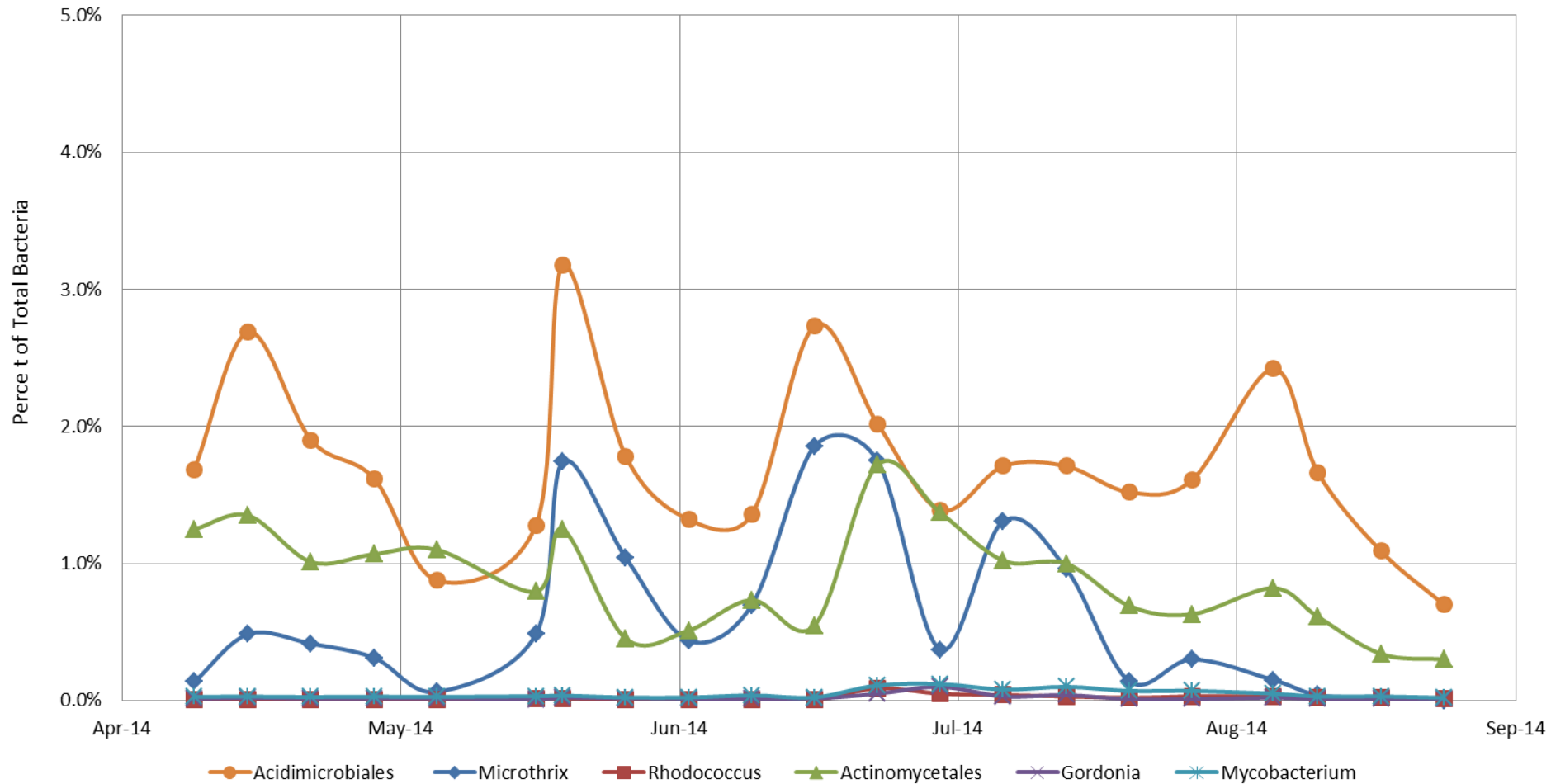


Filamentous Bacteria Results



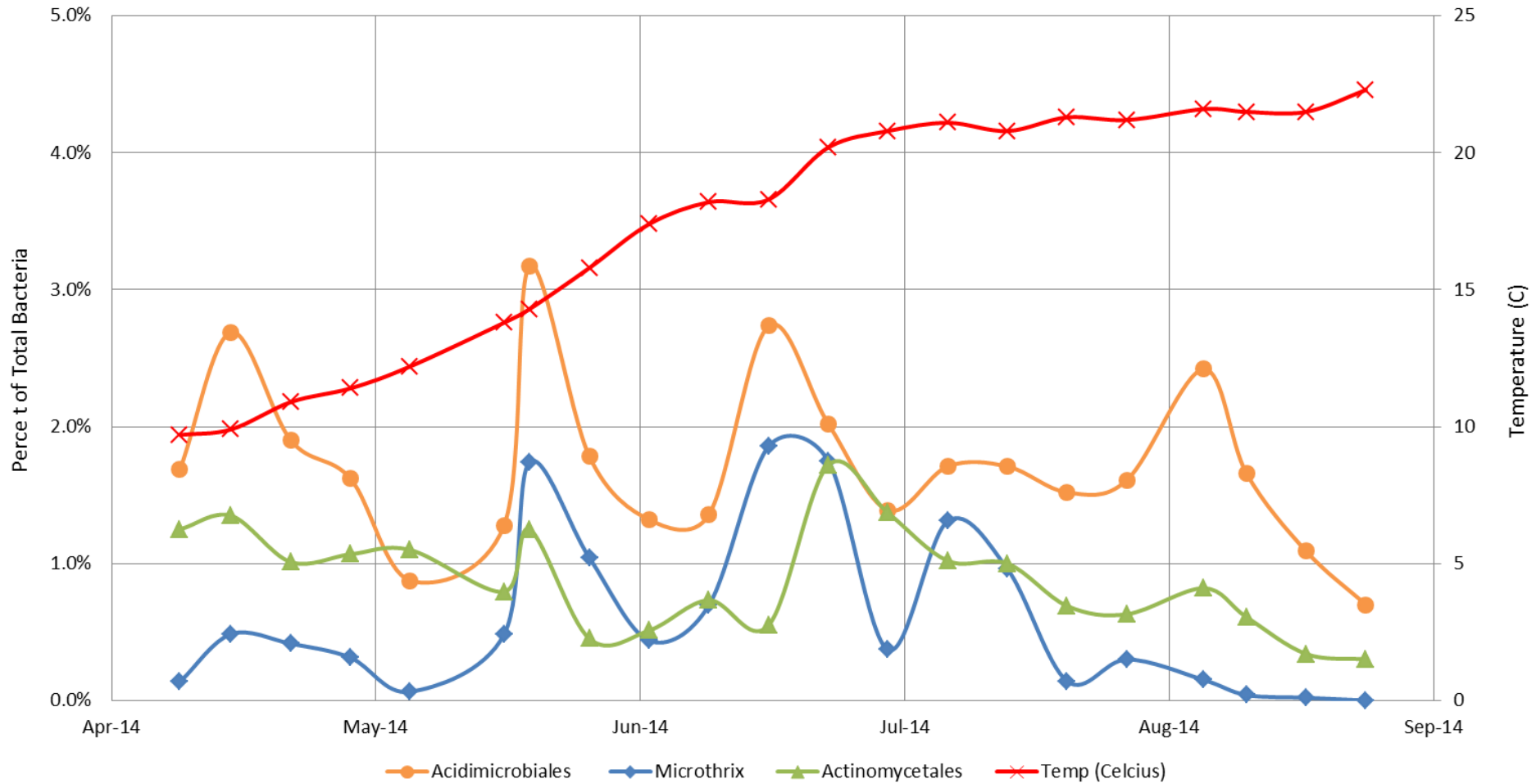
Foaming Bacteria Results

Foaming Bacteria



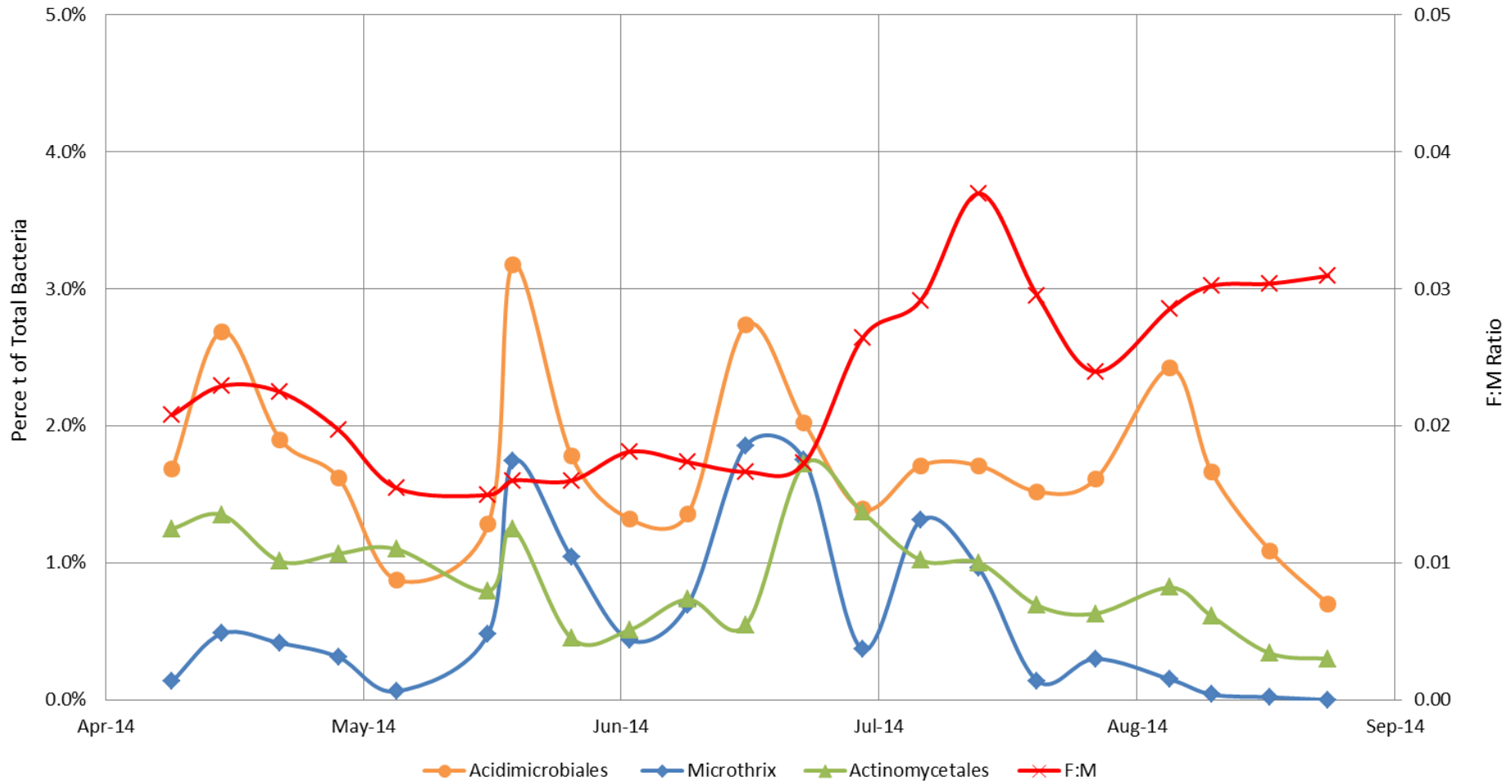
Foaming Bacteria Results

Foaming Bacteria vs Temperature



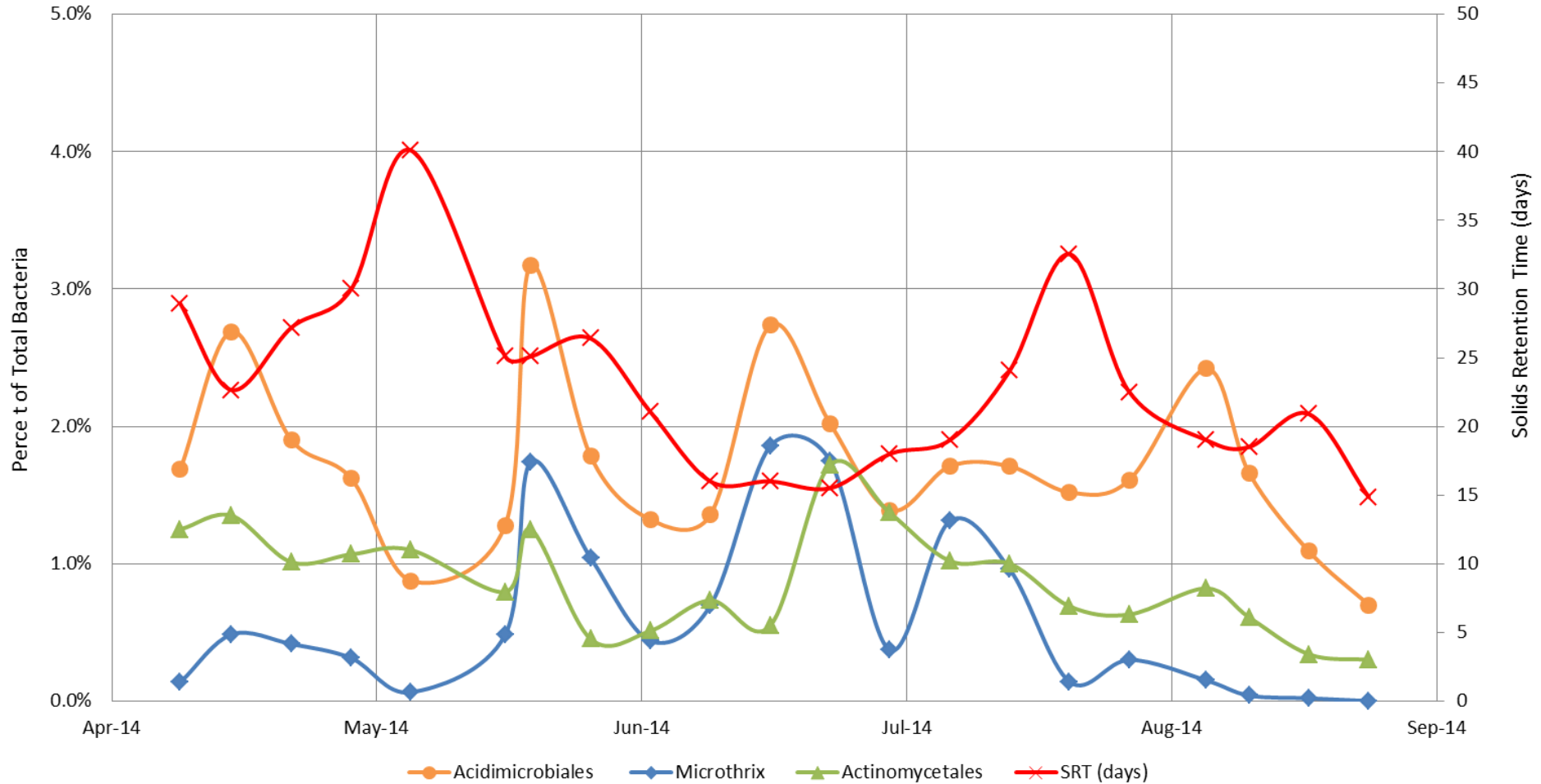
Foaming Bacteria Results

Foaming Bacteria with F:M Ratio



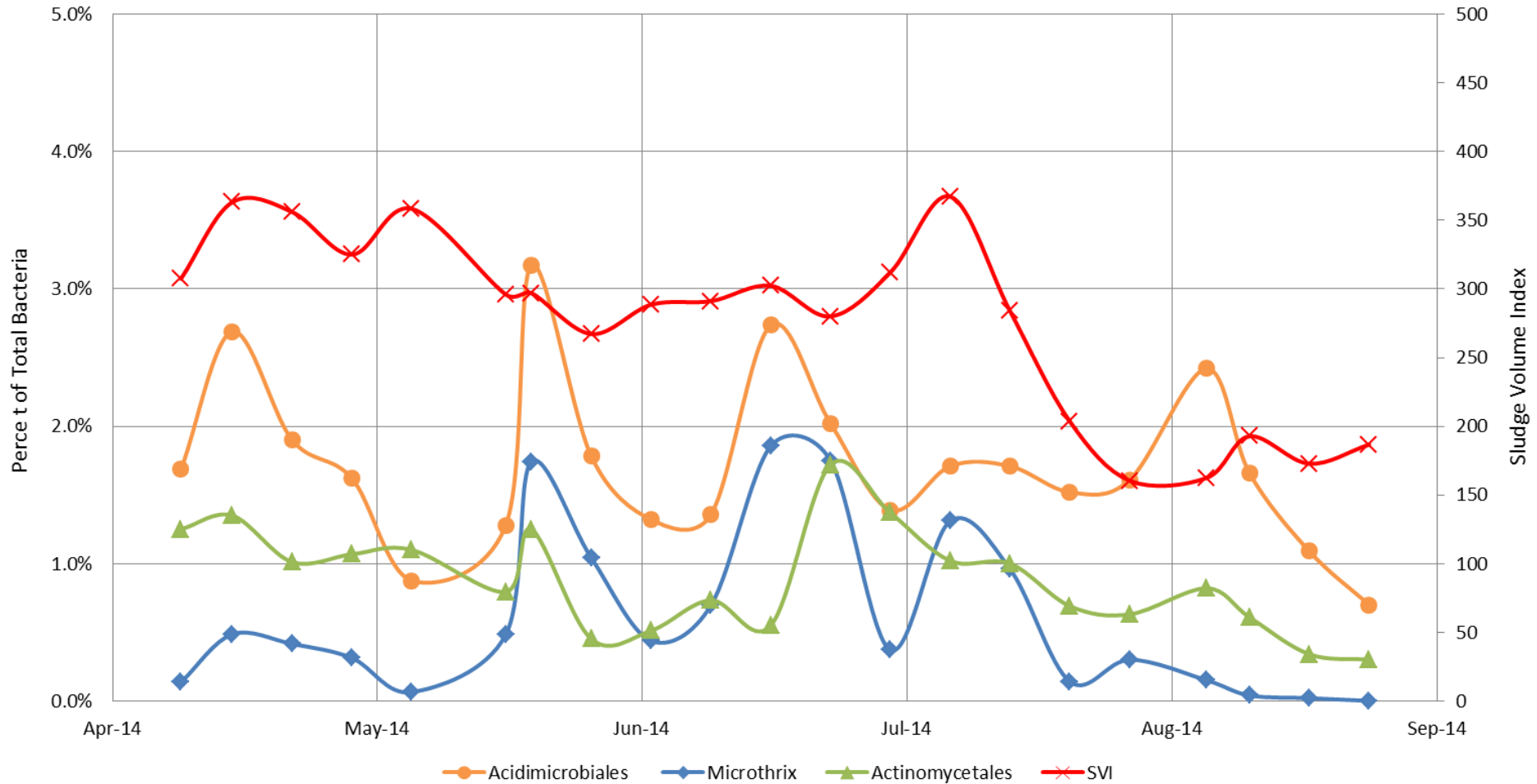
Foaming Bacteria Results

Foaming Bacteria vs SRT



Foaming Bacteria Results

Foaming Bacteria vs SVI



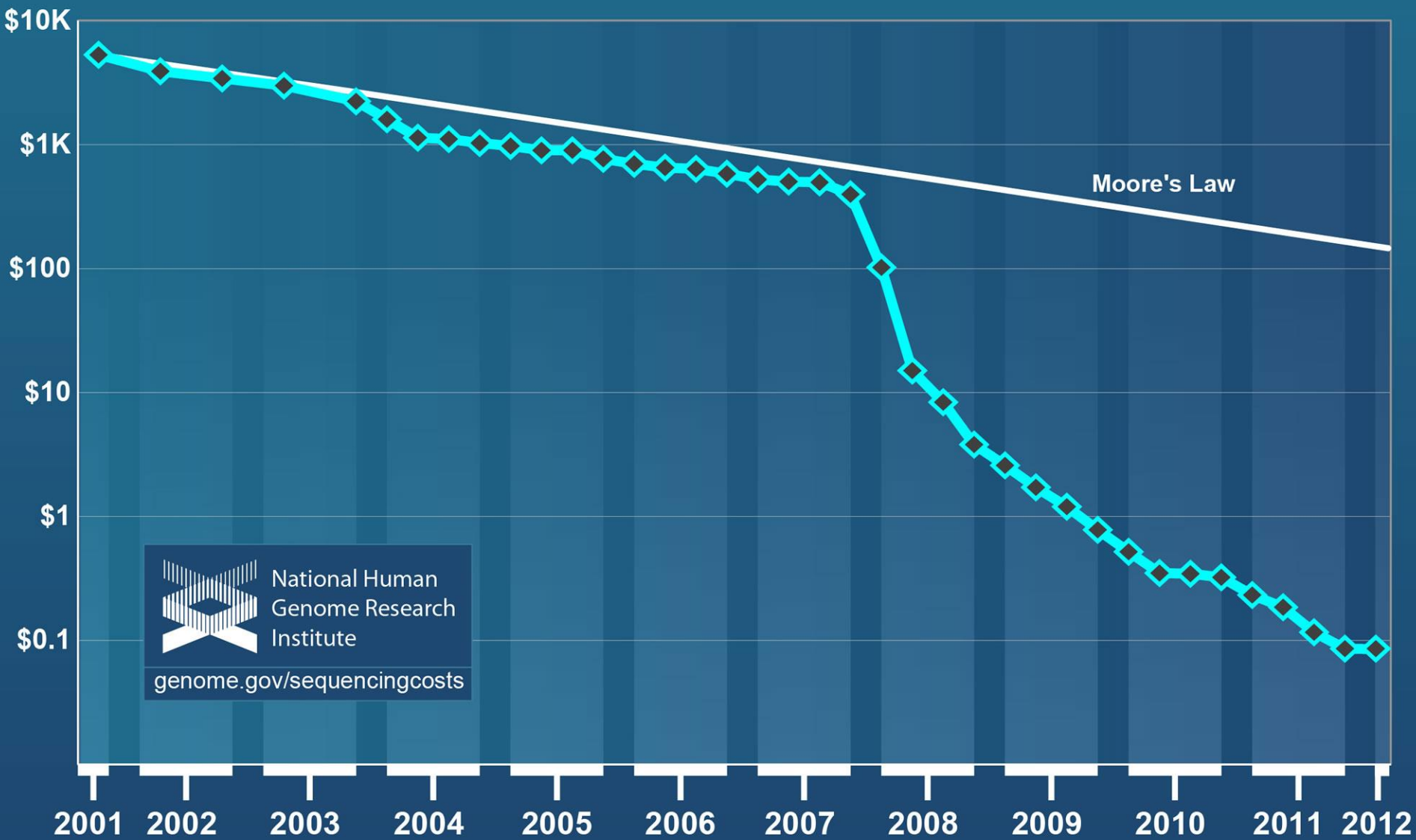
Results - Summary

- » Seasonal changes in Temperature, F:M, SRT caused bacteria population shift
 - Is the cause low temp, F:M, SRT?
- » Improved SVI (and effluent TSS) as Acidimicrobiales, Microthrix and Actinomyocetales decreased
- » What's up with Kouleothrix?

Cost

- » 21 tests = \$4,830 (\$230 per sample)
- » Cost increases with fewer samples

Cost per Raw Megabase of DNA Sequence



Conclusions

- » Bacteria culturing and microscopy have limitations
- » DNA speciation can help give operators clues to solving treatment process problems
- » DNA technology is an economical and effective method to assess entire microbial communities

Questions

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